

EXHIBIT B
PART 5 OF 5

in patients with chronic hepatitis C infections.^{18,19} Results of a phase I trial in patients with renal cell carcinoma demonstrated that there were no unique toxicities not seen with unmodified IFNs.²⁰

A second PEGylated interferon, IFN- α_{2b} , has also been developed. It is a conjugate with a 1:1 molar ratio of 12 kDa PEG.²¹ The pharmacokinetics and pharmacodynamics of this molecule indicate that it has a shorter half-life than Pegasys as predicted by the lower overall molecular weight.²² It has activity against CML, but has not been demonstrated to be statistically noninferior to nonPEGylated IFN- α_{2b} .²³ This conjugate also has activity to decrease platelet numbers in essential thrombocythemia.²⁴

The use of either PEGylated IFN- α in combination of ribavirin for treatment of HCV reduces the relative risk (RR) for remaining infected by 17% with nonPEGylated interferon plus ribavirin.²⁵ The combination treatment with the PEGylated proteins also results in a more cost effective approach to treating hepatitis C than the combination with unconjugated IFN- α .

3. PEGylated G-CSF

Granulocyte colony-stimulating factor is a member of the hematopoietic growth factor family that increases the numbers of neutrophilic granulocytes.² As such, G-CSF is an important supportive agent for cancer cytotoxic therapy. Although it appears to have no direct or indirect activity on cancer cells, its activity helps alleviate the neutrophilia associated with standard chemotherapy thereby allowing patients to tolerate higher doses of cytotoxic therapy. The so-called dose-dense chemotherapies, which use higher dosages than standard chemotherapy because of hematopoietic growth factor support, have shown promise in treatment of hematological and solid tumors, especially breast cancer.²⁶ In addition, G-CSF induces migration of hematopoietic stem cells from bone marrow into peripheral blood, making it possible to collect large numbers of stem cells by leukapheresis. This is a much more tolerable source of hematopoietic stem cells from donors, especially in the case of allogeneic transplants, than drawing bone marrow aspirates; although the same is true for obtaining these stem cells from patients who receive them back as autologous stem cell rescue.

Three different molecular forms of recombinant G-CSFs are registered for clinical use in various countries. Filgrastim is produced in *E. coli* and has an N-terminal methionine. Lenograstim is produced in Chinese hamster ovary cells and is therefore glycosylated.²⁷ Nartograstim is a mutein

produced in *E. coli* with five amino acids substitutions in the N-terminal portion that increase its specific activity.²⁸

A systematic approach to design a PEG-G-CSF that could be given once a week was undertaken. The goal was to make a molecule that would have pharmacokinetic properties that fit standard cancer chemotherapy practices that are usually designed on weekly schedules.

A number of different conjugates with one to three PEG chains of 5, 10, or 20 kDa per molecule of nartograstim were prepared. These conjugates were tested for their capacity to stimulate proliferation of G-CSF dependent cells *in vitro* and to increase granulocyte numbers in mice.²⁹ The activity *in vitro* was inversely proportional to the amount of PEG attached. The constructs having the highest the total molecular weight had the lowest the activity on cell proliferation in culture. This result suggests that the larger bulkier constructs may not bind as well to the G-CSF receptors. Interestingly, the *in vivo* activity was directly proportional to the total molecular weight of the constructs. Those molecules with the highest molecular weight had the highest activity in mice.²⁹ Similar results were evident in studies with cynomolgus monkeys where the higher molecular weight constructs had increased activities.³⁰ These results indicate that the duration of interaction between PEG-G-CSF and its receptors is more important than the affinity of this interaction.

A once weekly PEG-filgrastim (SD/01) has recently been produced based on the findings with PEG-nartograstim describe above.³¹ This molecule has a single PEG chain of 20 kDa molecular weight attached to the N-terminus of filgrastim.³² In mice, SD/01 causes a similar prolongation of neutrophilia in mice following a single injection and has been shown to reduce the mortality in an experimental murine model of invasive *Candida albicans* infection.³³ In clinical trials, a single administration of SD/01 results in prolonged elevation of neutrophils in patients receiving chemotherapy³⁴ as well as in normal volunteers.³¹ No dose limiting toxicities were seen. In the clinical studies, it was also demonstrated that the peripheral blood stem cell levels were elevated in the patients and volunteers to a comparable extent as with filgrastim injected daily at the same cumulative dose. A trial using a fixed dose of pegfilgrastim demonstrated equivalence to daily filgrastim and significantly reduced incidence of febrile neutropenia than found with filgrastim.³⁵

4. Other PEGylated Hematopoietic Growth Factors

A number of other hematopoietic growth factors have been PEGylated, but most published studies describe work in preclinical models. One factor that has similar activity to G-CSF is granulocyte macrophage colony-stimulating factor (GM-CSF). It has not received the same wide spread use as G-CSF because it is more toxic. The increased toxicity is due to the fact that it is, as the name implies, less lineage specific and stimulates macrophages as well as granulocytes. However, this additional activity may provide a rationale for its use in immunotherapy as an adjuvant for cancer vaccines. For vaccines to be effective, it is important to have the antigens presented in the proper manner to the immune effector cells. Dendritic cells, which are the most effective antigen presenting cells, can be expanded *in vitro* by GM-CSF.³⁶ A number of studies in animals have demonstrated that the high GM-CSF levels greatly enhance the antitumor effects of cancer vaccines.^{37,38} In a murine study, PEG-GM-CSF has been shown to expand a subpopulation of dendritic cells *in vivo*,³⁹ so could be an active adjuvant for cancer vaccines.

Another factor with clinical activity is megakaryocyte growth and development factor (MGDF). This is a truncated form of thrombopoietin. MGDF has been shown to enhance platelet recovery in mice following bone marrow transplantation and the PEGylated form has a single 20 kDa PEG chain attached to the amino terminus.⁴⁰ This modification helps stabilize the tertiary structure of the truncated protein and gives it approximately 3-fold more active than the unmodified version.⁴¹ The half life *in vivo* is short compared to that of PEG-filgrastim described above, so that that daily dosing is required. Two phase I trials have been reported, one in cancer patients receiving moderately myelosuppressive chemotherapy⁴² and the other in breast cancer patients receiving high-dose therapy with autologous peripheral blood stem cell support.⁴³ Both studies showed that the drug was well tolerated, even at the maximum doses tested of 5 and 10 $\mu\text{g}/\text{kg}/\text{d}$, respectively. A recent report,⁴⁴ demonstrated PEG-MGDF enhances platelet numbers in cancer patients receiving chemotherapy. Unfortunately, this molecule has not had a significant effect on platelet levels in patients receiving high dose therapy for leukemia or in those receiving stem cell support after chemotherapy.⁴⁰

PEG-MGDF as a single agent mobilizes stem cells, but did not significantly enhance mobilization above that achieved by chemotherapy plus filgrastim in these patients.

Interleukin-6 (IL-6) is another thrombopoietic factor that has been PEGylated.⁴⁵ Because its toxicity is greater than that of PEG-MGDF, it seems unlikely that it will play a significant role in clinical practice.

5. PEGylated Lymphokines

Another set of cytokines that have been used in cancer treatment trials are called lymphokines because they are produced by lymphocytes and often act on lymphocytes. One, Interleukin-2 (IL-2), formerly known as T-cell growth factor has been used in a number of clinical studies, but it has not become part of standard clinical practice because of its high toxicity. This toxicity has limited its use primarily to major academic medical centers where there is more experience dealing with these toxicities. A phase I clinical trial demonstrated that a PEGylated IL-2 conjugate with 10–20-fold increased half-life had similar toxicities to those of the unmodified protein.⁴⁶ Direct injection of PEG-IL-2 into locoregional recurrent head and neck tumors appears to be less toxic than systemic administration, resulting in only temporary local swelling and redness. The response rate was not sufficient to justify additional studies. Interestingly, the one tumor that responded was small.⁴⁷ Low dose PEG-IL-2 has been used in patients with HIV with relatively low toxicity and objective stimulation of the immune system.^{48,49} A potential therapeutic use for this molecule in cancer may be in minimal residual disease settings, or in combination with anticancer vaccines.

Tumor necrosis factor- α (TNF- α), as the name implies, has direct cytotoxic effects on tumor cells. Initial studies in mice suggested that the human factor had little effect on normal cells. However, this was due to lack of species cross-reactivity for one type of TNF receptor, but not another.⁵⁰ In fact, TNF- α has considerable toxicity in patients. Preclinical studies with PEG-TNF- α conjugates prepared with different molecular weight PEG chains demonstrated that there was an optimal molecular size of 100–110 kDa for *in vitro* activity in mice.⁵¹

PEGylation of IFN- γ , a type II interferon produced by lymphocytes, has been tested in animals and is reported to have a lower affinity for its receptor but higher *in vivo* activity in rats,⁵² which is similar to situation

with PEG-G-CSF. This IFN has not found wide clinical use because its toxicities are similar to those of IL-2 in relation to potential therapeutic benefit.

6. PEGylated Cytokine Inhibitors

In addition to PEGylation of cytokines for therapeutic use, there are also efforts to develop PEGylated protein inhibitors of cytokine action. Because cytokines have powerful effects on the immune system, such as those described above for IL-2 and TNF, inhibition of their activity may be beneficial for indications such as toxic shock and tumor cachexia as well as chronic conditions like arthritis, asthma, Crohn's disease and autoimmune encephalomyelitis. These inhibitors are of two types: (1) soluble cytokine receptors, some of which are produced by cells and appear to be physiological inhibitors; (2) antibodies to the cytokines or to their receptors.

Soluble receptors against TNF, IL-1, IL-4, IL-5, and IFN- γ have been studied in preclinical models and in some cases clinical trials.⁵³ A recombinant TNF receptor hybrid with the Fc portion of human immunoglobulin has been registered for use in rheumatoid arthritis. PEGylated TNF receptor constructs have been reported to be active in a variety of models^{54,55} and might be useful for this indication.

Antibody reagents can be directed against the cytokine or the receptor itself. A PEGylated antibody against IL-8 has been prepared with potential application in adult respiratory distress syndrome.⁵⁵ An important anti-cancer antibody, trastuzumab (HerceptinTM), is directed against the oncogene ERB-B2 (also called Her-2/Neu), a member of the epidermal growth factor receptor family that is overexpressed in about 30% of breast cancers as well as a number of other tumor types. Treatment of patients having ERB-B2 positive tumors with the humanized antibody trastuzumab leads to regression of the tumors, presumably because the antibody blocks the cell growth stimulus from the receptor and causes the cells to undergo programmed cell death (apoptosis). A PEGylated construct of Herceptin has been tested in animal models and has prolonged activity compared to the unmodified antibody.⁵⁶

7. Conclusions and Outlook

After decades of repeated cycles of promises and failures, immunotherapy has finally lead to effective treatments for cancer. At this time, these have

primarily come from antibody therapeutics such as trastuzumab and rituximab (Rituxan™), which is directed against the CD20 antigen expressed on some B-cell nonHodgkin's lymphomas.⁵⁷ The protein drugs that have the largest sales in the cancer market, however, are for support of standard toxic therapies, namely the hematopoietic growth factors.⁵⁸ Studies over the years with hematopoietic cells in culture have shown that the continuous presence of the protein is necessary for proliferation, differentiation, and even survival of these cells.⁵⁹⁻⁶¹ These factors all have short half-lives and thus, exogenous injection has limited effectiveness.

PEGylation can significantly extend the plasma half-lives these proteins, not only allowing less frequent administration, but also enhancing their efficacy. Until recently, most of the cytokine constructs had relatively small amounts of PEG attached so the increases in half-life were modest. Modifications of interferon and G-CSF with high molecular weight PEG have shown that such constructs can provide therapeutic benefits. One reason this approach was not taken previously was that constructs were initially selected on the basis of maintaining high activity in various *in vitro* assays. This can lead to selection against the most active forms *in vivo*. Therefore, following the experience with PEG-G-CSFs, it is likely that higher molecular weight constructs with decreased *in vitro* activities will be appearing in the future.

As our knowledge of the immune system increases, our ability to develop effective immunotherapies, including vaccines is enhanced. Hopefully the future will provide a range of agents that will help the body fight cancers as they form.

References

1. Pfeffer LM, Dinarello CA, Herberman RB, Williams BR, Borden EC, Bordens R, Walter MR, Nagabhushan TL, Trotta PP and Pestka S (1998) Biological properties of recombinant alpha-interferons: 40th anniversary of the discovery of interferons. *Cancer Res* 58:2489-2499.
2. Welte K, Gahrilove J, Bronchud MH, Platzer E and Morstyn G (1996) Filgrastim (r-metHuG-CSF): The first 10 years. *Blood* 88:1907-1929.
3. Mehvar R (2000) Modulation of the pharmacokinetics and pharmacodynamics of proteins by polyethylene glycol conjugation. *J Pharm Pharm Sci* 3:125-136.
4. Yamaoka T, Tabata Y and Ikada Y (1994) Distribution and tissue uptake of poly(ethylene glycol) with different molecular weights after intravenous administration to mice. *J Pharm Sci* 83:601-606.

5. Kuwabara T, Kobayashi S and Sugiyama Y (1996) Kinetic analysis of receptor-mediated endocytosis of G-CSF derivative, nartograstim, in rat bone marrow cells. *Am J Physiol* 271:E73–E84.
6. Tushinski RJ, Oliver IT, Guilbert LJ, Tynan PW, Warner JR and Stanley ER (1982) Survival of mononuclear phagocytes depends on a lineage-specific growth factor that the differentiated cells selectively destroy. *Cell* 28:71–81.
7. Veronese FM (2001) Peptide and protein PEGylation: A review of problems and solutions. *Biomaterials* 22:405–417.
8. Francis GE, Fisher D, Delgado C, Malik F, Gardiner A and Neale D (1998) PEGylation of cytokines and other therapeutic proteins and peptides: The importance of biological optimization of coupling techniques. *Int J Hematol* 68:1–18.
9. Keating MJ, Holmes R, Lerner S and Ho DH (1993) L-asparaginase and PEG asparaginase—past, present and future. *Leukemia Lymphoma* 10:153–157.
10. Ettinger LJ, Kurtzberg J, Voute PA, Jurgens H and Halpern SL (1995) An open-label, multicenter study of polyethylene glycol-L-asparaginase for the treatment of acute lymphoblastic leukemia. *Cancer* 75:1176–1181.
11. Ho DH, Brown NS, Yen A, Holmes R, Keating M, Abuchowski A, Newman RA and Krakoff IH (1986) Clinical pharmacology of polyethylene glycol-L-asparaginase. *Drug Metab Dispos* 14:349–352.
12. Gutterman JU (1994) Cytokine therapeutics — lessons from interferon alpha. *Proc Nat Acad Sci USA* 91:1198–1205.
13. Lippman SM, Parkinson DR, Itri LM, Weber RS, Schantz SP, Ota DM, Schusterman MA, Krakoff IH, Gutterman JU and Hong WK (1992) 13-cis-retinoic acid and interferon alpha-2a — effective combination therapy for advanced squamous cell carcinoma of the skin. *J Nat Cancer Inst* 84:235–241.
14. Lippman SM, Kavanagh JJ, Paredes-Espinoza M, Delgadillo-Madrueno F, Paredes-Casillas P, Hong WK, Holdener E and Krakoff IH (1992) 13-cis-retinoic acid plus interferon alpha-2a — highly active systemic therapy for squamous cell carcinoma of the cervix. *J Nat Cancer Inst* 84:241–245.
15. Monkarsh SP, Ma Y, Aglione A, Bailon P, Ciolek D, DeBarbieri B, Graves MC, Hollfelder K, Michel H, Palleroni A, Porter JE, Russoman E, Roy S and Pan YC (1997) Positional isomers of monopegylated interferon alpha-2a: Isolation, characterization and biological activity. *Anal Biochem* 247:434–440.
16. Nieforth KA, Nadeau R, Patel IH and Mould D (1996) Use of an indirect pharmacodynamic stimulation model of MX protein induction to compare *in vivo* activity of interferon alfa-2a and a polyethylene glycol-modified derivative in healthy subjects. *Clin Pharmacol Ther* 59:636–646.
17. Bailon P, Palleroni A, Schaffer CA, Spence CL, Fung WJ, Porter JE, Ehrlich GK, Pan W, Xu ZX, Modi MW, Farid A, Berthold W and Graves M (2001) Rational design of a potent, long-lasting form of interferon: A 40 kDa

- branched polyethylene glycol-conjugated interferon alpha-2a for the treatment of hepatitis C. *Bioconjug Chem* 12:195–202.
18. Heathcote EJ, Shiffman ML, Cooksley WG, Dusheiko GM, Lee SS, Balart L, Reindollar R, Reddy RK, Wright TL, Lin A, Hoffman J and De Pamphilis J (2000) Peginterferon alfa-2a in patients with chronic hepatitis C and cirrhosis. *N Engl J Med* 343:1673–1680.
 19. Zeuzem S, Feinman SV, Rasenack J, Heathcote EJ, Lai MY, Gane E, O'Grady J, Reichen J, Diago M, Lin A, Hoffman J and Brunda MJ (2000) Peginterferon alfa-2a in patients with chronic hepatitis C. *N Engl J Med* 343:1666–1672.
 20. Motzer RJ, Rakhit A, Ginsberg M, Rittweger K, Vuky J, Yu R, Fettner S and Hooftman L (2001) Phase I trial of 40-kd branched pegylated interferon alfa-2a for patients with advanced renal cell carcinoma. *J Clin Oncol* 19:1312–1319.
 21. Wang YS, Youngster S, Bausch J, Zhang R, McNemar C and Wyss DF (2000) Identification of the major positional isomer of pegylated interferon alpha-2b. *Biochemistry* 39:10634–10640.
 22. Glue P, Fang JW, Rouzier-Panis R, Raffanel C, Sabo R, Gupta SK, Salfi M and Jacobs S (2000) Pegylated interferon-alpha2b: Pharmacokinetics, pharmacodynamics, safety and preliminary efficacy data. Hepatitis C Intervention Therapy Group. *Clin Pharmacol Ther* 68:556–567.
 23. Michallet M, Maloisel F, Delain M, Hellmann A, Rosas A, Silver RT and Tendler C (2004) Pegylated recombinant interferon alpha-2b vs recombinant interferon alpha-2b for the initial treatment of chronic-phase chronic myelogenous leukemia: A phase III study. *Leukemia* 18:309–315.
 24. Alvarado Y, Cortes J, Verstovsek S, Thomas D, Faderl S, Estrov Z, Kantarjian H and Giles FJ (2003) Pilot study of pegylated interferon-alpha 2b in patients with essential thrombocythemia. *Cancer Chemother Pharmacol* 51:81–86.
 25. Shepherd J, Brodin H, Cave C, Waugh N, Price A and Gabbay J (2004) Pegylated interferon alpha-2a and -2b in combination with ribavirin in the treatment of chronic hepatitis C: A systematic review and economic evaluation. *Health Technol Assess* 8:iii–iv, 1–125.
 26. Citron ML, Berry DA, Cirrincione C, Hudis C, Winer EP, Gradishar WJ, Davidson NE, Martino S, Livingston R, Ingle JN, Perez EA, Carpenter J, Hurd D, Holland JF, Smith BL, Sartor CI, Leung EH, Abrams J, Schilsky RL, Muss HB and Norton L (2003) Randomized trial of dose-dense versus conventionally scheduled and sequential versus concurrent combination chemotherapy as postoperative adjuvant treatment of node-positive primary breast cancer: First report of intergroup trial C9741/cancer and leukemia group B trial 9741. *J Clin Oncol* 21:1431–1439.
 27. Baumann I, Testa NG, Lange C, Dewynter E, Luft T, Dexter TM, Vanhoef MEHM and Howell A (1993) Haemopoietic cells mobilized into the circulation

- by lenograstim as alternative to bone marrow for allogeneic transplants. *Lancet* **341**:369.
28. Kuga T, Komatsu Y, Yamasaki M, Sekine S, Miyaji H, Nishi T, Sato M, Yokoo Y, Asano M, Okabe M, Morimoto M and Itoh S (1989) Mutagenesis of human granulocyte colony stimulating factor. *Biochem Biophys Res Commun* **159**:103–111.
 29. Bowen S, Tare N, Inoue T, Yamasaki M, Okabe M, Horii I and Eliason JF (1999) Relationship between molecular mass and duration of activity of polyethylene glycol conjugated granulocyte colony-stimulating factor mutein. *Exp Hematol* **27**:425–432.
 30. Eliason JF, Greway A, Tare N, Inoue T, Bowen S, Dar M, Yamasaki M, Okabe M and Horii I (2000) Extended activity in cynomolgus monkeys of a granulocyte colony-stimulating factor mutein conjugated with high molecular weight polyethylene glycol. *Stem Cells* **18**:40–45.
 31. Molineux G, Kinstler O, Briddell B, Hartley C, McElroy P, Kerzic P, Sutherland W, Stoney G, Kern B, Fletcher FA, Cohen A, Korach E, Ulich T, McNiece I, Lockbaum P, Miller-Messana MA, Gardner S, Hunt T and Schwab G (1999) A new form of Filgrastim with sustained duration *in vivo* and enhanced ability to mobilize PBPC in both mice and humans. *Exp Hematol* **27**:1724–1734.
 32. Molineux G (2004) The design and development of pegfilgrastim (PEG-rmetHuG-CSF, Neulasta). *Curr Pharm Des* **10**:1235–1244.
 33. van Spriël AB, van den Herik-Oudijk IE and van de Winkel JG (2000) A single injection of polyethylene-glycol granulocyte colony-stimulating factor strongly prolongs survival of mice with systemic candidiasis. *Cytokine* **12**:666–670.
 34. Johnston E, Crawford J, Blackwell S, Bjurstrom T, Lockbaum P, Roskos L, Yang BB, Gardner S, Miller-Messana MA, Shoemaker D, Garst J and Schwab G (2000) Randomized, dose-escalation study of SD/01 compared with daily filgrastim in patients receiving chemotherapy. *J Clin Oncol* **18**:2522–2528.
 35. Green MD, Koelbl H, Baselga J, Galid A, Guillem V, Gascon P, Siena S, Lalisang RI, Samonigg H, Clemens MR, Zani V, Liang BC, Renwick J and Piccart MJ (2003) A randomized double-blind multicenter phase III study of fixed-dose single-administration pegfilgrastim versus daily filgrastim in patients receiving myelosuppressive chemotherapy. *Ann Oncol* **14**:29–35.
 36. Siena S, Di Nicola M, Bregni M, Mortarini R, Anichini A, Lombardi L, Ravagnani F, Parmiani G and Gianni AM (1995) Massive *ex vivo* generation of functional dendritic cells from mobilized CD34+ blood progenitors for anti-cancer therapy. *Exp Hematol* **23**:1463–1471.
 37. Golumbek PT (1993) Controlled release, biodegradable cytokine depots — a new approach in cancer vaccine design. *Cancer Res* **53**:5841–5844.

- therapy in patients with locoregionally recurrent head and neck squamous-cell carcinoma. *Ann Oncol* 5:957–960.
48. Tepler H, Kaplan G, Smith KA, Montana AL, Meyn P and Cohn ZA (1993) Prolonged immunostimulatory effect of low-dose polyethylene glycol interleukin 2 in patients with human immunodeficiency virus type 1 infection. *J Exp Med* 177:483–492.
 49. Ramachandran R, Katzenstein DA, Winters MA, Kundu SK and Merigan TC (1996) Polyethylene glycol-modified interleukin-2 and thymosin alpha 1 in human immunodeficiency virus type 1 infection. *J Infect Dis* 173:1005–1008.
 50. Lütscher H, Steinmetz M and Lesslauer W (1991) Tumor necrosis factor: Receptors and inhibitors. *Cancer Cells* 3:221–226.
 51. Tsutsumi Y, Kihira T, Tsunoda S, Kamada H, Nakagawa S, Kaneda Y, Kanamori T and Mayumi T (1996) Molecular design of hybrid tumor necrosis factor-alpha III: Polyethylene glycol-modified tumor necrosis factor-alpha has markedly enhanced antitumor potency due to longer plasma half-life and higher tumor accumulation. *J Pharmacol Exp Ther* 278:1006–1011.
 52. Kita Y, Rohde MF, Arakawa T, Fagin KD, Fish EN and Banerjee K (1990) Characterization of a polyethylene glycol conjugate of recombinant human interferon-gamma. *Drug Des Deliv* 6:157–167.
 53. Fernandez-Botran R (2000) Soluble cytokine receptors: Novel immunotherapeutic agents. *Expert Opin Invest Drugs* 9:497–514.
 54. Edwards CK (1999) PEGylated recombinant human soluble tumour necrosis factor receptor type I (r-Hu-sTNF-RI): novel high affinity TNF receptor designed for chronic inflammatory diseases. *Ann Rheum Dis* 58(Suppl 1): 173–181.
 55. Koumenis IL, Shahrokh Z, Leong S, Hsei V, Deforge L and Zapata G (2000) Modulating pharmacokinetics of an anti-interleukin-8 F(ab')₂ by amine-specific PEGylation with preserved bioactivity. *Int J Pharm* 198:83–95.
 56. Hurwitz E, Klapper LN, Wilchek M, Yarden Y and Sela M (2000) Inhibition of tumor growth by poly(ethylene glycol) derivatives of anti-ErbB2 antibodies. *Cancer Immunol Immunother* 49:226–234.
 57. Reichert J and Pavolu A (2004) Monoclonal antibodies market. *Nat Rev Drug Discov* 3:383–384.
 58. Pavlou AK and Reichert JM (2004) Recombinant protein therapeutics — success rates, market trends and values to 2010. *Nat Biotechnol* 22:1513–1519.
 59. Pélèraux A and Eliason JF (1989) Proliferation of single hemopoietic progenitor cells in the absence of colony-stimulating factors and serum. *Exp Hematol* 17:1032–1037.

60. Kan O, Baldwin SA and Whetton AD (1994) Apoptosis is regulated by the rate of glucose transport in an IL-3-dependent haemopoietic cell line. *Biochem Soc Trans* **22**:275S.
61. Metcalf D and Merchav S (1982) Effects of GM-CSF deprivation on precursors of granulocytes and macrophages. *J Cell Physiol* **112**:411-418.