

EXHIBIT I

Lodish Decl. in Support of Opposition to Roche's Motion for Summary Judgment of Invalidity for Double Patenting Over Claim 10 of the '016 Patent

Development and characterization of novel erythropoiesis stimulating protein (NESP)

Joan C. Egrie and Jeffrey K. Browne

Amgen Inc., 1 Amgen Center Drive, MS 27-4-A, Thousand Oaks, CA 91320-1799, USA

Abstract

Studies on human erythropoietin (EPO) demonstrated that there is a direct relationship between the sialic acid-containing carbohydrate content of the molecule and its serum half-life and *in vivo* biological activity, but an inverse relationship with its receptor binding affinity. These observations led to the hypothesis that increasing the carbohydrate content, beyond that found naturally, would lead to a molecule with enhanced biological activity. Hyperglycosylated recombinant human EPO (rHuEPO) analogues were developed to test this hypothesis. Darbepoetin alfa (novel erythropoiesis stimulating protein, NESP), which was engineered to contain five N-linked carbohydrate chains (two more than rHuEPO), has been evaluated in preclinical animal studies. Due to its increased sialic acid-containing carbohydrate content, NESP is biochemically distinct from rHuEPO, having an increased molecular weight and greater negative charge. Compared with rHuEPO, it has an approximately 3-fold longer serum half-life, greater *in vivo* potency, and can be administered less frequently to obtain the same biological response. NESP is currently being evaluated in human clinical trials for treatment of anaemia and reduction in its incidence.

Keywords: biological activity; carbohydrate; darbepoetin alfa; erythropoietin; pharmacokinetics; review

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Correspondence and offprint requests to: Dr Joan C. Egrie, Amgen Inc., 1 Amgen Center Drive, MS 27-4-A, Thousand Oaks, CA 91320-1799, USA.

Introduction

Erythropoietin (EPO) is a glycoprotein hormone that is the primary regulator of erythropoiesis, maintaining the body's red blood cell mass at an optimum level [1,2]. In response to a decrease in tissue oxygenation, EPO synthesis increases in the kidney. The secreted hormone binds to specific receptors on the surface of red blood cell precursors in the bone marrow, leading to their survival, proliferation and differentiation, and ultimately to an increase in haematocrit [3,4].

Since its introduction more than a decade ago, recombinant human EPO (rHuEPO) has become the standard of care in treating the anaemia associated with chronic renal failure (CRF). It is highly effective at correcting the anaemia, restoring energy levels, and increasing patient well-being and quality of life [5–8]. It has also been approved for the treatment of anaemia associated with cancer, HIV infection, and use in the surgical setting to decrease the need for allogeneic blood transfusions. For all indications, it has proved to be remarkably well tolerated and highly efficacious [9–11].

The recommended and usual therapy with rHuEPO is two to three times per week by subcutaneous or intravenous injection. For CRF patients, the duration of therapy is for the life of the patient, or until a successful kidney transplant restores kidney function, including the production of the natural hormone. For cancer patients, rHuEPO therapy is indicated for as long as the anaemia persists, generally through the entire course of chemotherapy.

It can be a hardship to administer rHuEPO two to three times per week, particularly for those patients who do not otherwise need to be seen in the clinic this frequently. In these cases, the patient needs to make a special trip to the clinic for his or her rHuEPO therapy. As is true for all growth factors, reduction in dose frequency results in a significant loss in efficiency. That is, the total weekly dose required when rHuEPO is administered once a week is greater than when it is administered as two to three divided

doses. It was anticipated that this clinical need could be addressed by creating a molecule with enhanced *in vivo* bioactivity to allow for less frequent dosing of patients.

To create a molecule with enhanced activity, research was initially directed towards elucidating those factors and structural features that control the *in vivo* activity of EPO [12]. This research led to the discovery and development of darbepoetin alfa, a novel erythropoiesis stimulating protein (NESP), that can be administered less frequently than epoetin [13].

Structure of EPO

Human EPO is a 30400 Da, heavily glycosylated protein hormone [14,15]. Sixty per cent (by weight) of the molecule is an invariant 165 amino acid single polypeptide chain containing two disulphide bonds [16,17]. The remaining 40% of the mass of the molecule is carbohydrate. Carbohydrate addition (glycosylation) is a post-translational event that results in the addition of sugar chains to specific asparagine (N-linked) or serine/threonine (O-linked) amino acids in the polypeptide. The carbohydrate portion of natural and recombinant human EPO consists of three N-linked sugar chains at Asn24, 38 and 83, and one O-linked (mucin type) sugar chain at Ser126 [18,19].

Structural determinations using nuclear magnetic resonance (NMR) spectroscopy [20] and X-ray crystallography [21] have indicated that human EPO is an elongated molecule with an overall topology of a left-handed four-helix bundle, typical of members of the haematopoietic growth factor family. In addition, these studies have identified the amino acids at the receptor binding sites. The carbohydrate addition sites are clustered at one end of the molecule, distal from the receptor binding site. While the four carbohydrate chains contribute ca. 40% of the mass of the hormone, they probably cover much of the surface of the molecule since they have an extended and flexible molecular structure.

In contrast to the invariant amino acid sequence of the protein portion of glycoproteins, the carbohydrate structures are variable, a feature referred to as microheterogeneity. For example, N-glycosylation sites on the same protein may contain different carbohydrate structures. Furthermore, even at the same glycosylation site on a given glycoprotein, different structures may be found. This heterogeneity is a consequence of the non-template-directed synthesis of carbohydrates.

The carbohydrate structures of EPO have been determined and the extent of the microheterogeneity defined for both rHuEPO and the natural hormone [22-25]. One of the most prominent examples of microheterogeneity for EPO is seen on the N-linked carbohydrate chains, where the oligosaccharides may contain two, three or four branches (or antennae), each of which is typically terminated with the negatively

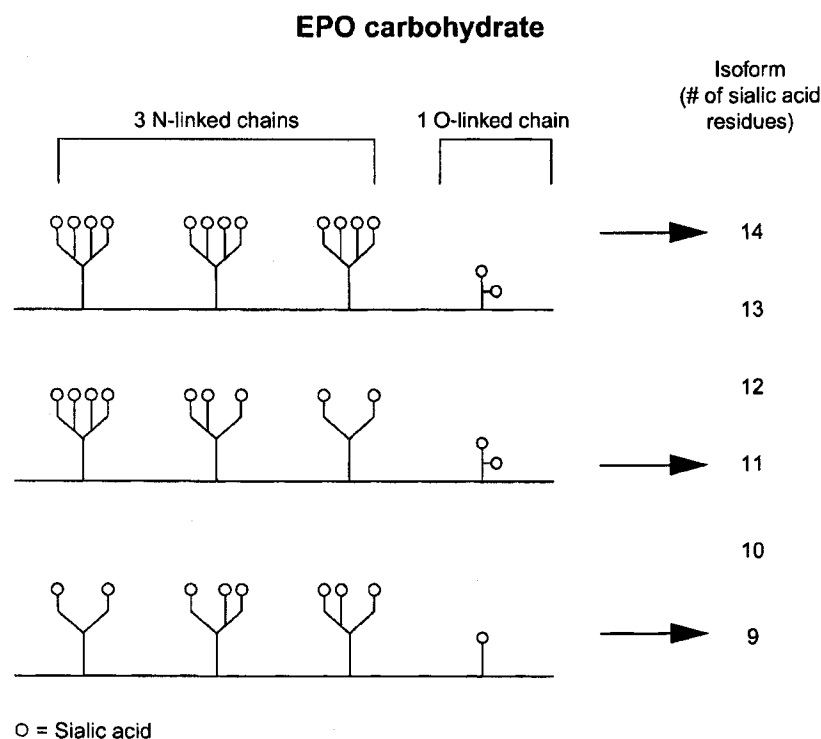
charged sugar molecule, sialic acid (Figure 1). With the exception of sialic acid, all of the other sugar molecules on EPO are neutral. Similarly, the single O-linked carbohydrate may contain zero to two sialic acid molecules. Since each of the three N-linked oligosaccharides can contain up to four sialic acid residues and the single O-linked chain can contain two, the EPO molecule can have a maximum of 14 sialic acid residues. Therefore, because of the variability in sugar structure, the number of sialic acid molecules on EPO varies and, as a consequence, so does the molecule's net negative charge. As indicated in Figure 1, an isoform of EPO is defined as a subset of the EPO molecules that has a defined charge due to its sialic acid content. For reference, epoetin alfa (Amgen Inc., Thousand Oaks, CA), the source of the purified rHuEPO used for these studies, has been purified so as to contain isoforms 9-14.

Role of carbohydrate in biological activity

The carbohydrate portions of different glycoprotein molecules have been shown to have many diverse functions, including effects on the biosynthesis and secretion, immune protection, conformation, stability, solubility and biological activity of molecules [26,27]. For rHuEPO, in particular, it has been shown that the addition of carbohydrate is required for secretion from the cell, and for increasing the solubility of the molecule [28-30]. Early research on EPO from natural sources indicated that the sialic acid residues were necessary for biological activity *in vivo* [31-33]. Removal of the sialic acid from either native EPO or rHuEPO resulted in molecules having an increased activity *in vitro*, but very low activity *in vivo*, presumably due to removal from circulation by the asialoglycoprotein receptor in the liver [34,35]. Similarly, it was shown that EPO molecules, which have been deglycosylated to remove carbohydrate (or produced in *Escherichia coli* to allow expression of only the EPO polypeptide), are active *in vitro*, but have very low *in vivo* activity [36,37].

In order to define further the role of carbohydrate in biological activity, the approach taken was to purify EPO carbohydrate isoforms, measure their *in vivo* activity and determine how the different carbohydrate structures affect activity.

rHuEPO, produced by Chinese hamster ovary cells, was purified to contain the entire complement of isoforms 4-14, and then fractionated further by ion exchange chromatography to isolate the individual isoforms [12]. The *in vivo* efficacy of each of the individual isoforms was tested in normal mice to determine the effect of repetitive dosing on the haematocrit. In this assay, CD-1 mice were injected with either a vehicle control or an equimolar dose (2.5 µg/kg of peptide) of each of the individual isoforms by intraperitoneal injection three times a week for 1 month. The results of this experiment



The structure of the carbohydrate chains is variable



Isolate molecules based on sialic acid content
Test the isolated EPO isoforms for biological activity

Fig. 1. Schematic of EPO carbohydrate structure and EPO isoform designation.

demonstrated a striking difference in the biological activity of the individual isoforms, with those isoforms having a higher sialic acid content exhibiting a progressively higher *in vivo* efficacy (Figure 2). By day 30, the group mean haematocrit of isoform 14-treated animals increased by 26.2 ± 2.7 points (to a haematocrit of 76.2%), compared with an increase of only 6.3 ± 3.5 points for the isoform 8-treated group; a 4.2-fold increase in efficacy. In contrast, animals receiving vehicle control showed no haematocrit change from baseline during the experiment.

It was reasoned that these results might have two possible explanations: the more active isoforms might have a longer serum half-life and/or an increased ability to bind to the EPO receptor. In order to assess the contribution of each of these possibilities, the pharmacokinetics and receptor binding activity of the individual isoforms was measured.

The isolated EPO isoforms were iodinated and their circulating half-life determined after intravenous injection into rats. At specified intervals after dosing, blood samples were taken and the fraction of iodinated isoform remaining in circulation was measured. The isoforms with the increased sialic acid content had a

longer serum half-life than those with the lower sialic acid content (Figure 3). The β -half-life of isoform 14 was 3.2-fold longer than that for isoform 6 (3.97 vs 1.24 h, respectively). As expected from these results, the serum clearance of the isoforms progressively increased as the sialic acid content decreased. In contrast, the volume of distribution was the same for the individual isoforms and approximately equal to the plasma volume (data not shown). Thus, those isoforms that have a higher sialic acid content have a higher *in vivo* biological activity, longer serum half-life and slower serum clearance.

Next, the relative affinity of various EPO isoform preparations for the EPO receptor was determined in a radioreceptor assay [38]. This assay measures the quantity of each isoform required to displace ^{125}I -rHuEPO bound to the EPO receptor on the surface of OCIM1 cells. The IC_{50} , the amount of test compound required to compete 50% of the receptor-bound ^{125}I -rHuEPO, was determined for each isoform. As seen in Figure 3, the higher the sialic acid content, the greater the quantity of EPO isoform necessary to compete ^{125}I -rHuEPO binding. Thus, those isoforms with a higher sialic acid content had a lower relative

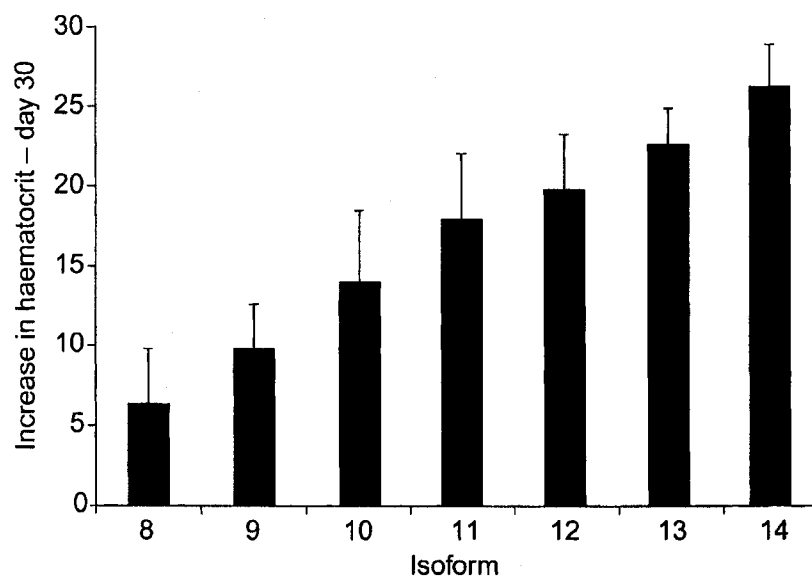


Fig. 2. *In vivo* efficacy of isolated EPO isoforms. CD-1 mice ($n=20$ /group) received an equimolar dose ($2.5 \mu\text{g}/\text{kg}$ of peptide) of each of the isolated EPO isoforms or vehicle control three times per week for 30 days by intraperitoneal injection. The haematocrit of each mouse was determined at baseline and twice weekly thereafter. At the conclusion of the study, final serum samples from all mice were screened for anti-rHuEPO antibodies by a radioimmunoprecipitation assay [19]. Mice that developed a significant anti-rHuEPO antibody response were excluded from the analysis. The increase in haematocrit over baseline following 30 days of treatment (day 30) was calculated for each mouse and the group average haematocrit increase (\pm SD) calculated for each group. There was no significant change in haematocrit in the vehicle control group over the course of the study ($46.1 \pm 1.2\%$ vs $45.2 \pm 1.2\%$ for baseline and day 30, respectively).

affinity for the EPO receptor. The relative affinity of isoform 6 for the EPO receptor was 7-fold greater than that for isoform 14.

Taken together, these experiments indicate that the carbohydrate moieties of EPO have significant effects on the biological activity of the hormone, modulating both receptor affinity and serum clearance. There is a direct relationship between sialic acid content, *in vivo* biological activity and serum half-life, but an inverse relationship with receptor affinity. While conventional wisdom might have predicted that increases in receptor affinity would lead to a more active molecule, clearly this is not the case. In fact, as shown in these experiments, isoform 9, which has a 2.6-fold greater affinity for the EPO receptor than isoform 14, has only approximately one-third of the *in vivo* activity (Figures 2 and 3). These observations clearly demonstrate that clearance has a far stronger influence on *in vivo* activity than receptor binding affinity. Increases in serum half-life were able to overcome the observed decreases in receptor affinity. Thus, for EPO, serum clearance rather than receptor binding affinity is the primary determinant of *in vivo* activity.

Design of hyperglycosylated rHuEPO analogues

In addition to identifying serum half-life as a major controlling factor of the *in vivo* biological activity of EPO, these experiments led to the hypothesis that increasing the sialic acid-containing carbohydrate of EPO, would increase its serum half-life and thereby the *in vivo* biological activity of the molecule.

To test this hypothesis, additional N-linked carbohydrate chains were added to the rHuEPO molecule. N-linked carbohydrate is attached to the polypeptide backbone at a consensus sequence for carbohydrate addition (Asn-Xxx-Ser/Thr). To introduce new carbohydrate attachment sites into the polypeptide backbone, the DNA sequence of the cloned human EPO gene needed to be modified to code for one or more new consensus sequences. The consensus sequences needed to be added at positions that were compatible with carbohydrate addition. While the consensus sequence is necessary for carbohydrate addition, it is not sufficient to ensure that a carbohydrate addition site will be utilized. Other factors, such as the local protein folding and conformation during biosynthesis, determine whether an oligosaccharide is attached at a given consensus sequence site. In addition, the consensus sequences needed to be added to positions that did not interfere with receptor binding, or compromise the folding, conformation or stability of the molecule.

At the time that these studies were initiated, there was only an incomplete understanding of the three-dimensional structure of EPO. This understanding was largely gained through site-directed mutagenesis studies where individual amino acids were changed, and the result of the change evaluated for effects on bioactivity and conformation [39,40]. These structure/function experiments identified many of the amino acids that were critical for EPO receptor interaction and necessary for proper folding of the molecule. Although these studies provided an insight as to which amino acids should not be altered, they did not allow identification of

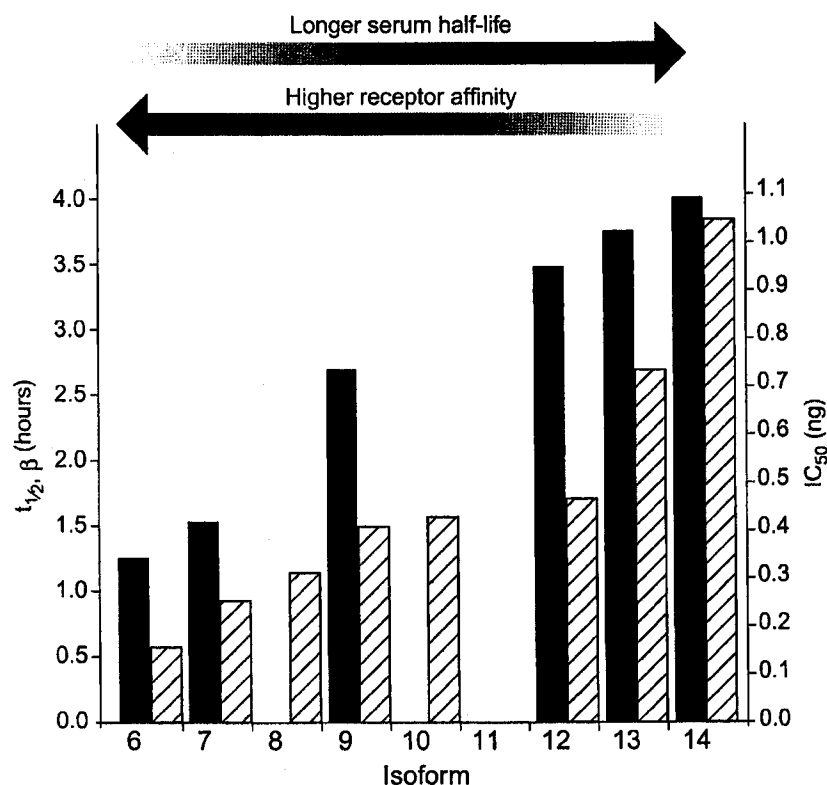


Fig. 3. EPO isoforms having a higher sialic acid content have a longer serum half-life and a lower receptor binding activity. The serum half-life (solid bars) and receptor binding activity (hatched bars) were determined for various isolated EPO isoform preparations. To determine serum half-life, iodinated isoforms were administered intravenously into cannulated Sprague-Dawley rats ($n=4/\text{group}$). Blood samples were collected and the fraction of ethanol-precipitable iodinated isoform remaining in the serum was measured. The receptor binding activity of isolated EPO isoforms was measured using a radioreceptor assay [38]. Increasing concentrations of each unlabelled isolated EPO isoform were incubated with 0.5 ng of ^{125}I -rHuEPO (a mixture of isoforms 9–14) and OCIM1 cells. Cell receptor-bound ^{125}I -rHuEPO was measured following separation from unbound ^{125}I -rHuEPO. Linear regression analysis was used to calculate the IC_{50} (the concentration of unlabelled EPO isoform required to compete 50% of the amount of ^{125}I -rHuEPO bound in the absence of cold competitor). The IC_{50} for unlabelled rHuEPO was 0.54 ng.

positions where extra carbohydrate addition sites could be added whilst maintaining structure.

To test this hypothesis, several dozen analogues of rHuEPO containing one or more amino acid substitutions, which created one or more new carbohydrate addition sites, were produced [41]. Site-directed mutagenesis was first used to change the nucleic acid sequence encoding one or more amino acids of a human EPO cDNA clone. Next, the clone encoding each new candidate analogue was transfected into mammalian cells and the expressed protein analysed. Only a few of the several dozen analogues tested were fully glycosylated, had the proper tertiary structure and retained biological activity. The EPO analogues that were properly glycosylated each had one extra N-linked carbohydrate chain (4-chain analogue). By combining the carbohydrate addition sites of two successfully glycosylated 4-chain analogues into one molecule, the 5-N-linked chain analogue, NESP, was created. The amino acid sequence of NESP differs from that of human EPO at five positions (Ala30Asn, His32Thr, Pro87Val, Trp88Asn and Pro90Thr), allowing for additional oligosaccharide attachment at asparagine residues at positions 30 and 88 [41].

Due to the additional carbohydrate, these 4- and 5-N-linked (NESP) chain analogues are each biochemically distinct from rHuEPO (Figure 4). They have an increased molecular weight, sialic acid content and negative charge. Each additional N-linked carbohydrate chain increases the molecular weight of the protein by ca. 3300 Da and adds up to four additional sialic acid residues. Thus, in comparison with rHuEPO, the two extra carbohydrate chains on NESP increase the molecular weight by 22% (to 37100 Da) and the maximum number of sialic acid residues from 14 to 22.

Biological activity of hyperglycosylated rHuEPO analogues

These analogues, having 4- and 5-N-linked carbohydrate chains, were used to test the hypothesis that increasing the sialic acid-containing carbohydrate content of EPO would increase the serum half-life and thereby the *in vivo* bioactivity.

The *in vivo* efficacy of rHuEPO, the 4-chain analogue and NESP (the 5-chain analogue) were

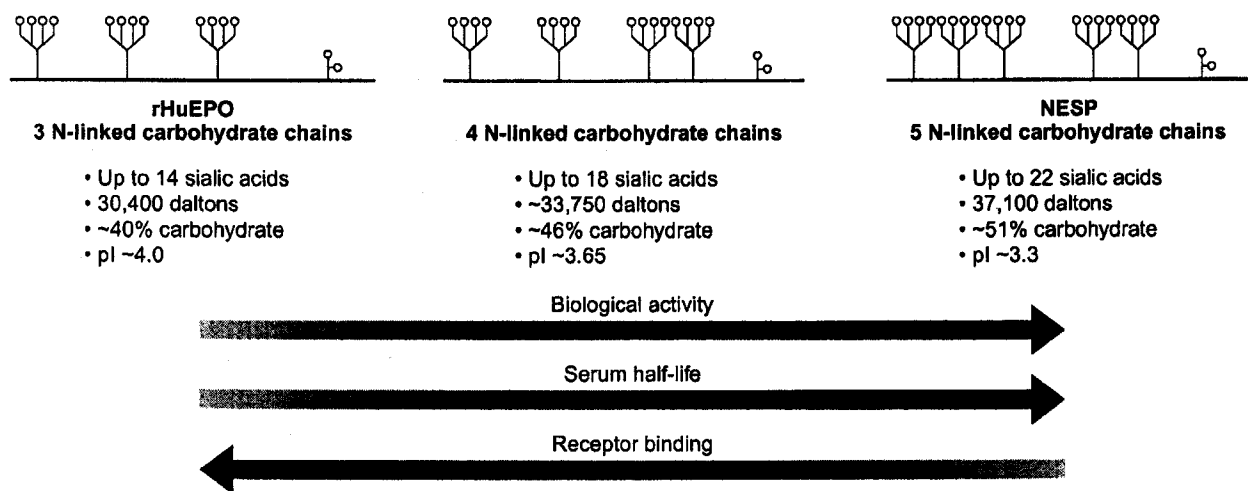


Fig. 4. Biochemical and biological properties of rHuEPO and rHuEPO analogues containing 4- and 5-N-linked carbohydrate chains [13].

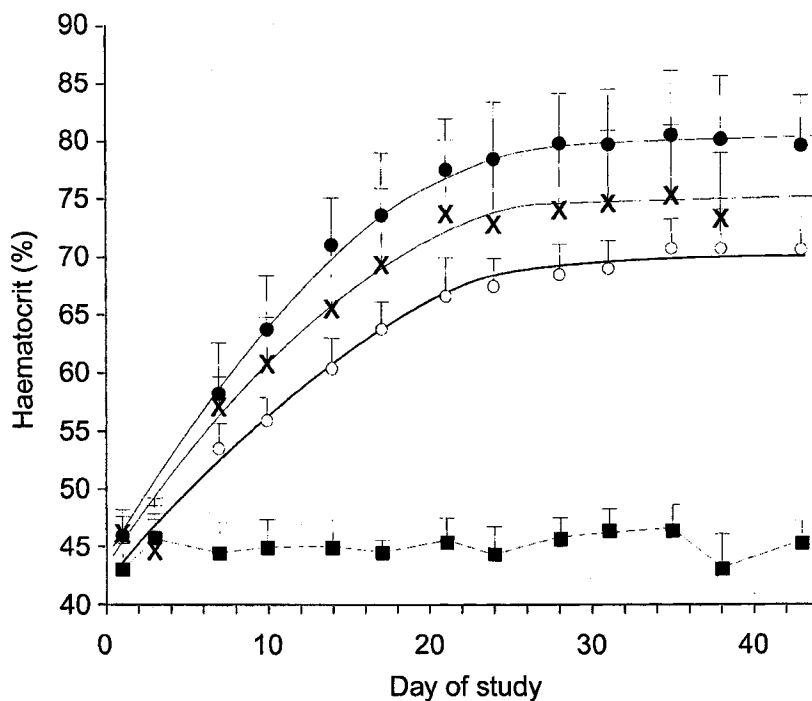


Fig. 5. *In vivo* efficacy of rHuEPO (isoforms 9–14), 4-chain analogue, and NESP. CD-1 mice (8–13/group) received an equimolar dose (2.5 µg/kg of peptide) of rHuEPO (O), 4-chain analogue (X), NESP (●) or vehicle control (■), three times per week for 42 days by intraperitoneal injection. Each point represents the group average haematocrit (±SD) for anti-rHuEPO antibody-negative mice (see Figure 2).

compared by measuring the increase in haematocrit of mice injected thrice weekly with equimolar doses (2.5 µg/kg of peptide) of each molecule for 6 weeks (Figure 5). Both NESP and the 4-chain analogue produced a faster rate of haematocrit rise and a higher stable plateau haematocrit than rHuEPO. By day 31 the haematocrit had increased by 22.8 ± 2.3 , 28.1 ± 6.8 and 33.9 ± 3.4 points for mice treated with rHuEPO, 4-chain analogue, and NESP, respectively, and these haematocrit differences were maintained for the duration of the experiment. Consistent with the

hypothesis, the magnitude of the haematocrit increase correlated with the number of carbohydrate chains and the sialic acid content of the molecules. Thus, by day 31, treatment with the 5-chain analogue, NESP, increased the haematocrit 11 points more than rHuEPO, while the response produced by the 4-chain analogue was intermediate, between that of NESP and rHuEPO.

To confirm that the mechanism of increased activity was due to an increase in the circulating half-life, the pharmacokinetics of rHuEPO, the 4-chain analogue,

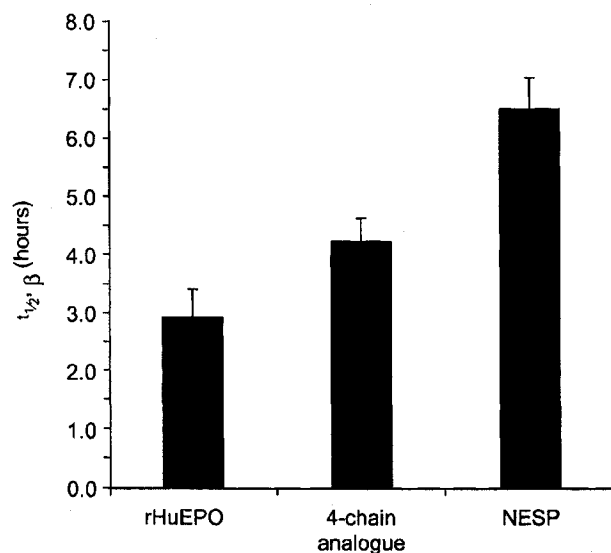


Fig. 6. Serum half-life of rHuEPO (isoforms 9–14), 4-chain analogue, and NESP. The test molecules were iodinated and administered intravenously into cannulated Sprague-Dawley rats ($n=10$ for rHuEPO, $n=6$ /group for 4-chain analogue, and NESP). Blood samples were collected periodically and the amount of ethanol-precipitable iodinated test compound remaining in the serum was measured.

and NESP were compared. As seen in Figure 6, the greater the sialic acid-containing carbohydrate content of the molecule, the longer its circulating half-life. The β -half-life of both hyperglycosylated molecules was greater than that determined for rHuEPO and for isoform 14, the highest of the naturally occurring EPO isoforms. In contrast, the volume of distribution was equivalent for the three molecules and approximately equal to the plasma volume (data not shown).

rHuEPO, the 4-chain analogue, and NESP were tested for EPO receptor binding activity in the radio-receptor assay. Consistent with the results obtained in this assay for the isolated isoforms and the hypothesis, the relative EPO receptor binding affinity was inversely correlated with the carbohydrate content. The relative affinity of NESP for the EPO receptor was 4.3-fold lower than that of rHuEPO and significantly lower than that of isoform 14. The results with the 4-chain analogue were intermediate between those of NESP and rHuEPO (data not shown).

These results confirm the hypothesis that the serum half-life of rHuEPO could be extended by increasing the sialic acid-containing carbohydrate content beyond that found naturally, and that the longer serum half-life would lead to an increase in *in vivo* biological activity (Figure 4). The results with the 4- and 5-chain hyperglycosylated EPO analogues were also consistent with the previous observations that the increases in serum half-life more than offset the decreases in receptor binding affinity. The *in vivo* biological activity of both the 4-chain analogue, and NESP were greater than that of rHuEPO, even though they each had a lower affinity for the EPO receptor.

Comparison of the *in vivo* efficacy of NESP with rHuEPO

To further characterize NESP and to evaluate its potential as a therapeutic for human use, comparative pharmacodynamic studies of NESP and rHuEPO were performed in normal mice using different frequencies and routes of administration over wide dose ranges. In particular, these studies focused on determining if the increase in serum half-life and *in vivo* activity would confer the clinical benefit of allowing for less frequent dosing.

As has been shown for rHuEPO, NESP produced a dose-dependent increase in the haematocrit of normal mice when injected by the intravenous, intraperitoneal and subcutaneous routes [13]. Initial experiments focused on comparing the efficacy of the two molecules when administered three times per week. In the experiment shown in Figure 7, thrice weekly intravenous dosing with 1.25 $\mu\text{g}/\text{kg}$ of NESP increased the haematocrit of mice to ca. 75% in 6 weeks; however, a comparable haematocrit increase with rHuEPO at this dosing frequency required 5.0 $\mu\text{g}/\text{kg}$. When the data from all experiments were combined, NESP was determined to be ca. 3.6-fold more potent than rHuEPO when administered three times a week by any route.

NESP can successfully increase the haematocrit of mice when administered once a week (Figure 7). A once weekly dose of 15 $\mu\text{g}/\text{kg}$ NESP produced a nearly identical biological response in normal mice, as did a thrice weekly dose of 1.25 $\mu\text{g}/\text{kg}$ (3.75 $\mu\text{g}/\text{kg}$ total weekly dose). Thus, for NESP, an approximate 4-fold weekly dose increase is required to change from thrice to once weekly dosing in this animal model. It is important to note, however, that in human clinical studies, there were no apparent differences between once weekly and thrice weekly dosing with NESP [42]. The optimal weekly dose was the same whether administered once a week or as three divided doses. The differences observed between the small animal model and human clinical trials are presumably due to the differences in the erythrokinetics and red blood cell lifespan of the two species.

Once weekly dosing with rHuEPO is far less efficient than once weekly dosing with NESP. As shown in Figure 7, once weekly dosing with 22.5 $\mu\text{g}/\text{kg}$ rHuEPO increased the haematocrit by seven points in 6 weeks, while a 2.5 $\mu\text{g}/\text{kg}$ dose administered three times per week (7.5 $\mu\text{g}/\text{kg}$ total weekly dose) increased the haematocrit by 22 points in the same time period. Integration of the data from all experiments demonstrated that for rHuEPO, a ca. 15-fold weekly dose increase is required to change from thrice weekly to once weekly dosing in this animal model.

When the efficacy of once weekly dosing of NESP and rHuEPO were compared, a striking difference in the potency of the two molecules was observed. As seen in Figure 7, a 3.75 $\mu\text{g}/\text{kg}$ dose of NESP administered once a week increased the haematocrit by

12 points. In contrast, a 6-fold higher dose of rHuEPO (22.5 $\mu\text{g}/\text{kg}$) increased the haematocrit by only seven points. When relative potency plots were constructed from all of the data, NESP was found to be 13- to 14-fold more potent than rHuEPO when each was administered once weekly.

As a consequence of the relative potency differences described above, NESP administered once weekly is as effective as the same total weekly dose of rHuEPO given as three divided doses (Figure 8). These experiments demonstrate that the same dose of NESP can be administered less frequently than rHuEPO to obtain

the same biological response. Furthermore, dosing with NESP as infrequently as once every other week can still increase the haematocrit of normal mice (data not shown).

As expected from the pharmacokinetic differences between NESP and rHuEPO, no one number can be used to express the relative potency difference between the two molecules. The relative potency of NESP and rHuEPO will necessarily change as a function of the dosing interval. Longer intervals between the administration of doses will lead to a greater potency difference. Thus, when each molecule is administered

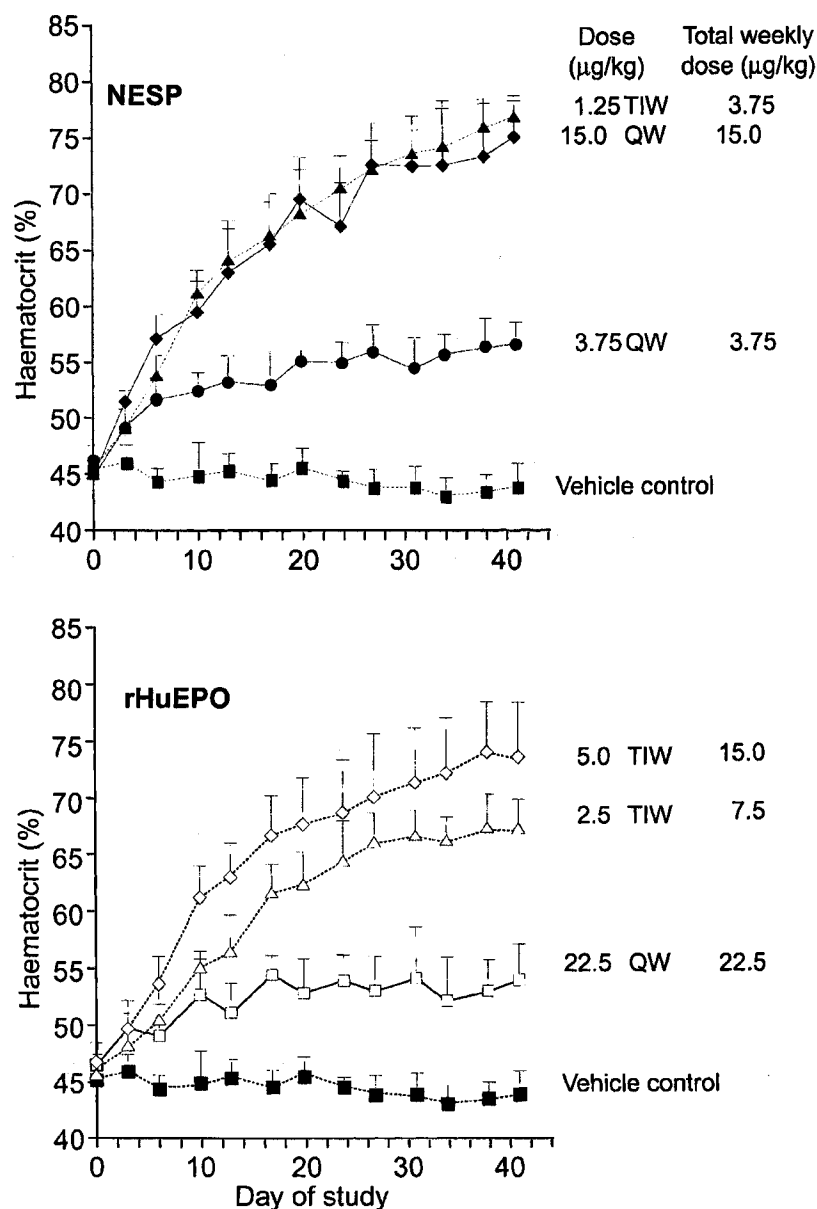


Fig. 7. *In vivo* efficacy of NESP and rHuEPO administered three times (TIW) or once (QW) a week. CD-1 mice ($n=7/\text{group}$) received rHuEPO (open symbols), NESP (closed symbols) or vehicle control (■) three times per week (TIW, dashed lines) or once a week (QW, solid lines) at the indicated dose levels for 42 days by intravenous injection. Each point represents the group average haematocrit (\pm SD) for the anti-rHuEPO antibody-negative mice (see Figure 2). Both the dose/administration and the total weekly dose are indicated on the graph.

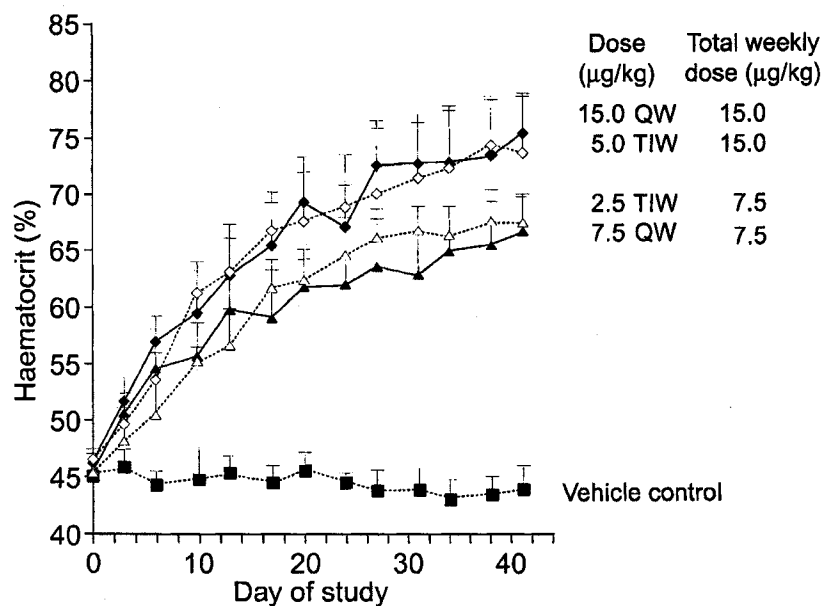


Fig. 8. Comparative efficacy of NESP administered once a week (QW) with rHuEPO administered three times per week (TIW). CD-1 mice ($n=7$ /group) received rHuEPO (open symbols) three times per week (TIW, dashed lines), NESP (closed symbols) once a week (QW, solid line) or vehicle control (■) once a week at the indicated dose levels for 42 days by intravenous injection. Each point represents the group average haematocrit (\pm SD) for the anti-rHuEPO antibody-negative mice (see Figure 2). Both the dose/administration and the total weekly dose are indicated on the graph.

three times per week, NESP is ca. 3.6-fold more potent than rHuEPO; however, when each molecule is administered once weekly, NESP is 13- to 14-fold more potent.

Comparative pharmacokinetics of NESP and epoetin alfa in dialysis patients

The first step in the clinical programme to determine whether NESP is both more potent than epoetin alfa and can be administered less frequently, was to verify that NESP had a longer serum half-life in patients. In a double-blind, randomized cross-over study, the single-dose pharmacokinetics of epoetin alfa (100 IU/kg) and an equimolar dose of NESP were compared after intravenous administration to 11 stable peritoneal dialysis patients [43]. Serum levels of NESP and rHuEPO were determined at regular intervals up to 96 h after dosing by immunoassay. In all patients, NESP had a longer terminal half-life than epoetin alfa (Figure 9). The mean terminal half-life for NESP following intravenous injection was 26.3 h, ca. 3-fold longer than that determined for epoetin alfa (8.5 h). There was no significant difference in the volume of distribution for the two molecules. This first study in man confirmed and extended the observations in animal studies with NESP, demonstrating that due to its increased sialic acid-containing carbohydrate content, NESP has a decreased clearance and longer serum half-life than rHuEPO.

Ongoing clinical studies are evaluating the safety and relative efficacy of NESP as compared with

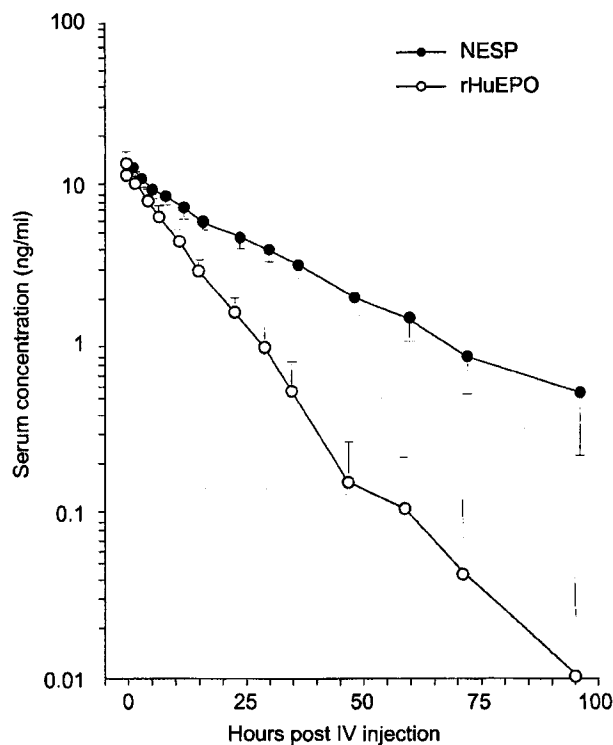


Fig. 9. Comparative pharmacokinetics of NESP and epoetin alfa in anaemic dialysis patients. Results are expressed as the mean (\pm SD). Reproduced by the kind permission of Lippincott Williams and Wilkins from Macdougall *et al.* 1999 [43].

epoetin alfa in different patient groups. Since the amino acid sequence of NESP differs at five amino acid positions from EPO, it is theoretically possible that NESP could be immunogenic. If NESP antibodies were to develop, they could be non-neutralizing (benign), or neutralizing, which would render NESP ineffective. In addition, either the neutralizing or non-neutralizing antibodies may cross-react with EPO. Due to this theoretical but real concern, patients are being closely monitored for the development of NESP antibodies in all clinical studies.

However, several important features of NESP suggest that the risk of immunogenicity may be minimal. One of the known functions of carbohydrate in glycoproteins is that it acts as a molecular shield, protecting the underlying polypeptide from the immune system [26]. Since the location of the five amino acid differences between NESP and EPO is at, or proximal to, the carbohydrate addition site, it is likely that these sites will be well shielded from immune surveillance. The fact that the new carbohydrate addition sites are distal to the receptor binding site minimizes the possibility that any antibodies that develop would be neutralizing. Carbohydrate chains in themselves are rarely immunogenic, and since all of the oligosaccharide structures on NESP are also found on epoetin, it is very unlikely that they will be immunogenic. Results from preclinical studies provide support for these considerations. In a mouse model, where the injection of heterologous human proteins is expected to lead to antibody formation, the incidence of seroconversion in NESP-treated animals was no higher than for rHuEPO-treated animals. Significantly, in all of the clinical trials to date, no patient has developed antibodies to NESP.

Conclusions

NESP has been engineered to contain five N-linked carbohydrate chains (two more than rHuEPO). The additional carbohydrate affects the biochemical and biological properties of NESP (Figure 4). Due to the additional sialic acid-containing carbohydrate, NESP has a slower serum clearance and, therefore, a longer half-life than rHuEPO. The longer serum half-life increases the *in vivo* biological activity and allows NESP to be administered less frequently than rHuEPO. The safety and efficacy of NESP for use as a therapeutic for the treatment of anaemia, and reduction in its incidence, is being evaluated in ongoing clinical trials.

The development of NESP is an outgrowth of basic research directed towards elucidating those structural features that control the *in vivo* biological activity of EPO. It was discovered that the pharmacokinetic properties of rHuEPO have a stronger influence on *in vivo* activity than receptor affinity and that the serum clearance of rHuEPO could be manipulated by changing the proportion of sialic acid-containing carbohydrate. These observations may be applicable for optimization of other protein therapeutics for clinical use.

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