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TABLE XI  
ECEPO SECTION 2

ECORI    KpnI  
A ATTCGGTACC AGAAC~~CC~~AAAG GTAACCTCT ACGCTGGAA ACCTA~~GGAA~~  
GCCATGG TCCTGGTTC CAATTTGAGA TGCGAACCCT TGCATACCTT

5                 AGGAGTGAAG AGGTGGCAG GGICGGCAC GGICGGCAC GGICGGCAC  
CAACCAGGT TCCGCACT TCAACCT CCAGACCCG ACGGATGGCT  
6                 GGICGAAC AGGGATGGCT CAAGCCGG CAAGCCGG CAAGCCGG  
CCGTAGAAC CACCGGTC CGTCGCCA TTAGGGGA GTGGCA  
10                 GGICGAAC AGGGATGGCT CAAGCCGG CAAGCCGG CAAGCCGG  
11                 GGICGAAC AGGGATGGCT CAAGCCGG CAAGCCGG CAAGCCGG  
12                 GGICGAAC AGGGATGGCT CAAGCCGG CAAGCCGG CAAGCCGG  
13                 AACGGIGCA GCGGCAGT GTAAGGACAG GAAGGACAG GAAGGACAG  
14                 GGACGGACGT CGACGTACAG CGTGTGC CATAGACGGGA CTCAGACCTAC

BamHI

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TABLE XII

ECEPO SECTION 3

1.	GATCCAGATCTCTGACTACTCTGC
5	2. ACGCAGCAGACTAGTCAGAGATCTG
	3. TCCGTGCTCTGGGTGCCACAGAAAGAGG
	4. GATAAACCTTTCTGTGCACCCAGAGC
	5. CTATCTCTCCGCCGGATGCTGCATCT
	6. CAGCAGATGCAGCATCCCCGGAGA
10	7. GCTGCACCGCTGCCTACCATCACTG
	8. ATCAGCAGTGATGCTACGGCAGCGCTG
	9. CTGATAACCTTCCGCAAACCTGTTTCG
	10. ATACACGAAACAGTTGCCGAAGGT
	11. TGTATACTCTAACTTCCCTGCCCTGGTA
15	12. CAGTTTACCCGGCTGGAAACTTAGAGT
	13. AACTGAAACTGTATACTGGCGAAGC
	14. CGCATGCTTCGCCAGTATACTAGTT
	15. ATGCCGTACTGGTGACCCCTAATAG
	16. TCGACTATTAGCGGTACCCAGTAC

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TABLE XIII  
ECEPO SECTION 3

BamHI BglII  
GA TCCAGATCTCTG  
GTCTAGAGAC

5

ACTACTCTGC 1 TCGCGTCTCT 3 GGGTGCACAG AAACAGGTTA 5 TCTCTCCGCC  
TGATGAGACG ACGCAGAGA CCCACGTGTC TTTCTCCGAT AGAGAGGCCGG  
2 4

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GGATGCTGCA TCTGCTGAC 6 CGGTGCGTAC CATCACTGT 7 GATACTTCC  
CCTACGACGT AGACGAATG GCGACCCATG GTAGTGACGA CTATGGAAGG  
6 8

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GCAAACTGTT TCGTGATATAC TCTAACTTCC 11 TGGGTGGTAA 13 ACTGAAACTG  
CGTTTGACAA AGCACATATG AGATTGAAGG ACGCACCATTT TGACTTTGAC  
10 12

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TATACTGGCG AACCATGCCG TACTGGTCAC CGCTAATAG 15 SalI  
ATATGACCGC TTCGTACGGC ATGACCACTG GCGATTATC AGCT  
14 16

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TABLE XIV  
ECEPO GENE

XbaI                          -1    1  
CTAG AAACCATGAG GGTAAATAAAA TAATGGCTCC GCGGGGTCTG  
TTGGTACTC CCATTATTT ATTACCGAGG CGGGGGAGAC  
5  
ATCTGCCACT CGAGAGTTCT GGAACGTTAC CTGCTGGAAG CTAAAGAAGC  
TAGACGCTGA GCTCTCAAGA CCTTGCAATG GACGACCTTC GATTCTTCG  
TGAAAACATC ACCACTGGTT GTGCTGAACA CTGTTCTTTG AACGAAAACA  
ACTTTGTAG TGGTGACCAA CACGACTTGT GACAAGAAC TTGCTTTGT  
10 TTACGGTACCC AGACACCAAG GTTAACCTCT ACGCTTGGAA ACGTATGGAA  
AATGCCATGG TCTGTGGTTC CAATTGAAGA TGCGAACCTT TGCTACCTT  
ATTGGTCAAC AAGCAAGTGA AGTTTGGCAG GGTCTGGCAC TGCTGAGCGA  
CAACCACTTG TTCGTCAACT TCAAACCGTC CCAGACCGTG ACGACTCGCT  
15 GGCTGTTACTG CGTGGCCAGG CACTGCTGGT AAACCTCTCT CAGCCGTGGG  
CCGACATGAC CCACCCGTCC CTGACGGACCA TTGAGGGAGA CCGGCACCC  
AACCCTGCTCCA CCTGCATGT GACAAAGCAG TATCTGGCT GAGATCTCTG  
TTGGCGACGT CGACGTACAA CTGTTTGTCA ATAGACCGGA CTCTAGAGAC  
ACTACTCTGC TGCCTGGCTCT GGGTGCACAG AAAGAGGCTA TCTCTCCGCC  
TGATGAGAGC ACGCACCGAGA CCCACGTGTC TTTCCTCGAT AGAGAGGGCG  
20 CGATGCTGCA TCTGCTGCCAC CGCTGCGTAC CATCACTGCT GATACTTCC  
CCTACGACGT AGACGACGTG GCGACGGCATG GTAGTGACGA CTATCCAAGG  
GCAAACTGTT CGGTGTATAAC TCTAACTTCC TCCGTTGGTAA ACTGAAACTG  
CGTTTGACAA AGCACATATG AGATTGAAGG ACGCACCATG TGACTTTGAC  
Sall  
TATACTGGCG AAGCATGCCG TACTGGTGAC CGCTAATAG  
ATATGACCGC TTGGTACGGC ATGACCACTG GCGATTATCA GCT  
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*Figure 10*

More particularly, Table VIII illustrates oligonucleotides employed to generate the Section 1 of the ECEPO gene encoding amino terminal residues of the human species polypeptide. Oligonucleotides were assembled into duplexes (1 and 2, 3 and 4, etc.) and the duplexes were then ligated to provide ECEPO Section 1 as *Figure 11*. Note that the assembled section includes respective terminal EcoRI and BamHI sticky ends, that "downstream" of the EcoRI sticky end is a XbaI restriction enzyme recognition site; and that "upstream" of the BamHI sticky end is a KpnI recognition site. Section 1 could readily be amplified using the M13 phage vector employed for verification of sequence of the section. Some difficulties were encountered in isolating the section as an XbaI/KpnI fragment from RF DNA generated in *E.coli*, likely due to methylation of the KpnI recognition site bases within the host. Single-stranded phage DNA was therefore isolated and rendered into double-stranded form *in vitro* by primer extension and the desired double-stranded fragment was thereafter readily isolated.

ECEPO gene Sections 2 and 3 (*Figures 13 and 15*) were constructed in a similar manner from the oligonucleotides of *Figures 12 and 14*, respectively. Each section was amplified in the M13 vector employed for sequence verification, and was isolated from phage DNA. As is apparent from *Table XI*, ECEPO Section 2 was constructed with EcoRI and BamHI sticky ends and could be isolated as a *KpnI/BglII* fragment. Similarly, ECEPO Section 3 was prepared with BamHI and SalI sticky ends and could be isolated from phage RF DNA as a *BglII/SalI* fragment. The three sections thus prepared can readily be assembled into a continuous DNA sequence (*Table XII*) encoding the entire human species EPO polypeptide with an amino terminal methionine codon (ATG) for *E.coli* translation initiation. Note also that "upstream" of the initial ATG is a series of base pairs substantially

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duplicating the ribosome binding site sequence of the highly expressed OMP-f gene of E.coli.

Any suitable expression vector may be employed to carry the ECEPO. The particular vector chosen for expression of the ECEPO gene as the "temperature sensitive" plasmid pCFM536 -- a derivative of plasmid pCFM414 (A.T.C.C. 40076) -- as described in co-pending U.S. Patent Application Serial No. 636,727, filed August 6, 1984, (Published EPo Application No 116,440) by Charles F. Morris. More specifically, pCFM536 was digested with XbaI and HindIII; the large fragment was isolated and employed in a two-part ligation with the ECEPO gene. Sections 1 (XbaI/KpnI), 2 (KpnI/BglII) and 3 (BglII/SalI) had previously been assembled in the correct order in M13 and the EPO gene was isolated therefrom as a single XbaI/HindIII fragment. This fragment included a portion of the polylinker from M13 mp9 phage spanning the SalI to HindIII sites therein. Control of expression in the resulting expression plasmid, p536, was by means of a lambda P<sub>L</sub> promoter, which itself may be under control of the C<sub>1857</sub> repressor gene (such as provided in E.coli strain K12ΔMtrp).

The manufactured ECEPO gene above may be variously modified to encode erythropoietin analogs such as [Asn<sup>2</sup>, des-Pro<sup>2</sup> through Ile<sup>6</sup>]hEPO and [His<sup>7</sup>]hEPO, as described below.

A. [Asn<sup>2</sup>, des-Pro<sup>2</sup> through Ile<sup>6</sup>]hEPO

Plasmid 536 carrying the ECEPO manufactured gene of Table XIV as a XbaI to HindIII insert was digested with HindIII and XbaI. The latter endonuclease cuts the ECEPO gene at a unique, 6 base pair recognition site spanning the last base of the codon encoding Asp<sup>8</sup> through the second base of the Arg<sup>10</sup> codon. A XbaI/XbaI "linker" sequence was manufactured having the following sequence:

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T0610X

<u>XbaI</u>	+1	2	7	8	9	<u>XhoI</u>
5'-CTAG	ATG	GCT	AAT	TGC	GAC-3'	
3'-TAC	CGA	TTA	ACG	CTG	AGCT-5'	

---

The XbaI/XhoI linker and the XhoI/HindIII ECEPO gene sequence fragment were inserted into the large fragment resulting from XbaI and HindIII digestion of plasmid pCFM526 -- a derivative of plasmid pCFM414 (A.T.C.C. 40076) -- as described in co-pending U.S. Patent Application Serial No. 636,727, filed August 6, 1984, by Charles F. Morris, to generate a plasmid-borne DNA sequence encoding E.coli expression of the Met<sup>-1</sup> form of the desired analog.

B. [His<sup>7</sup>]nEPO

Plasmid 536 was digested with HindIII and XhoI as in part A above. A XbaI/XhoI linker was manufactured having the following sequence:

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T0611X

<u>XbaI</u>	+1	2	3	4	5	6	7	8	9	<u>XhoI</u>
Met	Ala	Pro	Pro	Arg	Leu	Ile	His	Asp		
5'-CTAG	ATG	GCT	CCG	CCA	CGT	CTG	ATC	CAT	GAC-3'	
3'-TAC	CGA	GGC	GGT	GCA	GAC	TAG	GTA	CTG	AGCT-5'	

---

The linker and the XhoI/HindIII ECEPO sequence fragment were then inserted into pCFM526 to generate a plasmid-borne DNA sequence encoding E.coli expression of the Met<sup>-1</sup> form of the desired analog.

*(Handwritten notes: "No sense" with a checkmark, "BX", and "S" with a checkmark)*

Construction of a manufactured gene ("SCEPO") incorporating yeast preference codons is as described in the following Tables XV through XX. As was the case with the ECEPO gene, the entire construction involved formation of three sets of oligonucleotides (Tables XV and XX) which were formed into duplexes and assembled into sections (Tables XVI, XVII, XVIII and XX). Note that synthesis was facilitated in part by use of some sub-optimal codons in both the SCEPO and ECEPO construc-

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lations, i.e., oligonucleotides 7-12 of Section 1 of both genes were identical, as were oligonucleotides 1-6 of Section 2 in each gene.

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TABLE XV  
SCEPO SECTION 1 OLIGONUCLEOTIDES

1. AATTCAAGCTTGGATAAAAGAGCT
- 5 2. GTGGAGCTCTTTATCCAAGCTTG
3. CCACCAAGATTGATCTGTGACTC
4. TCTCGAGTCACAGATCAA~~C~~TTG
5. GAGAGTTTGGAAAGATA~~T~~TTGTTG
6. CTTCCAACAAGTATCTT~~T~~CCAAAAC
- 10 7. GAAGCTAAAGAAGCTGAAAACATC
8. GTGGTGTGTTTCAGCTTCTTAG
9. ACCACTGGTTGTGCTGAACACTGTT
10. CAAAGAACAGTGT~~T~~CAGCAC~~A~~CCA
11. TTTCAACGAAAAACATTACGGTACCG
- 15 12. GATCCGGTACCGTAATGTTTCGTT

TABLE XVI  
SCEPO SECTION 1

EcoRI    HindIII 1  
 20    AATTCA AGCTTGGATA  
       GT TCGAACCTAT  
       2

3  
 AAAGAGCTC ACCAAGATTG ATCTGTGACT C~~G~~AGAGTTT  
 TTTCTCGAGG T~~G~~T~~T~~CTAAC TAGACACTGA GCTCT~~C~~AAAAA  
       4

5  
 25    GGAAAGATA~~C~~ TTGTT~~G~~AAG CTAAAGA~~G~~C TGAAAACATC ACCACTGGTT  
       CCTTTCTAT~~S~~ AACAAAC~~S~~T~~C~~ GATT~~T~~CTTCG ACTTTTG~~T~~AG TGGT~~G~~ACCAA  
       6                      7  
       6                      8

9  
 30    GTGCTGAACA CTGTT~~T~~T~~T~~G AACGAA~~A~~ACA TTACGGTACC G~~C~~ KpnI    BamHI  
       CACGACTTG~~T~~ GACAAGAAAC~~T~~ TTGCTTTGT AATGCCATGG CCTAG  
       11                    12

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TABLE XVIISCEPO SECTION 2 OLIGONUCLEOTIDES

- |    |   |
|----|---|
| 1. | AATTCGGTACCCAGACACCAA <del>GCT</del>                  |
| 5  | 2. GTTAACCTTGGTGTCTGGTACCG                            |
|    | 3. TAACTTCTACGCTTGGAA <del>ACG</del> STAT             |
|    | 4. TTCCATACGTTCCAAGCGTAGAA                            |
|    | 5. GGAAGTTGGTCAACA <del>G</del> CAGTTGAAGT            |
|    | 6. CCAAACTTCAACTGC <del>T</del> TGTTGACCAAC           |
| 10 | 7. TTGGCAAGGT <del>T</del> TG <del>C</del> TTGTTATCTG |
|    | 8. GCTTCAGATAACAAGGCCAACCTG                           |
|    | 9. AAGCTGTTTGACAGGTCAAGCCT                            |
|    | 10. AACAAAGGCT <del>T</del> GACCTCTCAAAACA            |
|    | 11. TGTGGTTAACTCTTCTCAACCATGGG                        |
| 15 | 12. TGGTTCCCATGGTTGAGAAGAGTTAAC                       |
|    | 13. AACCATTGCAATTGCACGTGAT                            |
|    | 14. CTTTATGACGTGCAATTGCAA                             |
|    | 15. AAAGCCGTCTCTGGTTGAGATCTG                          |
|    | 16. GATCCAGATCTCAAAACCAGAGACGG                        |

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